

Synthesis, Electrochemistry, and Electrogenerated Chemiluminescence of Two BODIPY-Appended Bipyridine Homologues

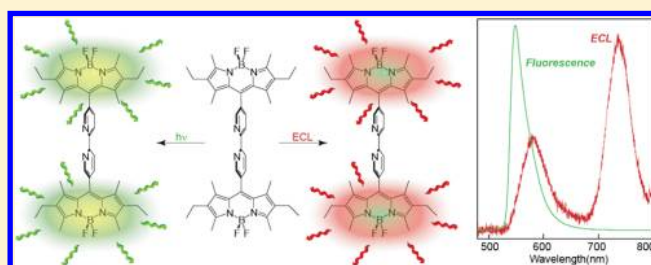
Honglan Qi,^{†,§} Justin J. Teesdale,[‡] Rachel C. Pupillo,[‡] Joel Rosenthal,^{*,‡} and Allen J. Bard^{*,†}

[†]Center for Electrochemistry, Department of Chemistry and Biochemistry, The University of Texas, Austin, Texas 78712, United States

[‡]Department of Chemistry and Biochemistry, University of Delaware, Newark, Delaware 19716, United States

S Supporting Information

ABSTRACT: Two new 2,2'-bipyridine (bpy) derivatives containing ancillary BODIPY chromophores attached at the 5- and 5'-positions (**BB3**) or 6- and 6'-positions (**BB4**) were prepared and characterized. In this work, the basic photophysics, electrochemistry, and electrogenerated chemiluminescence (ECL) of **BB3** and **BB4** are compared with those previously reported for a related bpy-BODIPY derivative (**BB2**) (*J. Phys. Chem. C* **2011**, *115*, 17993–18001). Cyclic voltammetry revealed that **BB3** and **BB4** display reversible 2e⁻ oxidation and reduction waves, which consist of two closely spaced (50–70 mV) 1e⁻ events. This redox behavior is consistent with the frontier molecular orbitals calculated for **BB3** and **BB4** and indicates that the 2,2'-bipyridine spacer of each bpy-BODIPY homologue does not facilitate efficient electronic communication between the tethered indacene units. In the presence of a coreactant such as tri-*n*-propylamine (TPA) or benzoyl peroxide (BPO), **BB3** and **BB4** exhibit strong ECL and produce spectra that are very similar to their corresponding photoluminescence profiles. The ECL signal obtained under annihilation conditions, however, is significantly different and is characterized by two distinct bands. One of these bands is centered at ~570 nm and is attributed to emission via an S- or T-route. The second band occurs at longer wavelengths and is centered around ~740 nm. The shape and concentration dependence of this long-wavelength ECL signal is not indicative of emission from an excimer or aggregate, but rather it suggests that a new emissive species is formed from the bpy-BODIPY luminophores during the annihilation process.



INTRODUCTION

Electrogenerated chemiluminescence (ECL) involves the generation of oxidized and reduced species, often radical ions, that undergo a fast electron transfer reaction to produce a species in an excited state.¹ The various mechanisms by which ECL processes take place have been extensively discussed.^{2–5} A prime pathway by which ECL is achieved is radical ion annihilation. An alternative mechanism involves consumption of a coreactant, which generates a reductant upon oxidation (or oxidant upon reduction) that reacts with the emission precursor.⁶ Coreactants can often be used to help elucidate ECL reaction mechanisms by providing a contrasting pathway to that observed for annihilation ECL.^{7,8} ECL is often produced by directly generating an excited singlet state (S-route) in which the annihilation energy is sufficient to populate the singlet excited state. Alternatively in cases where this energy is not sufficient to populate the singlet state, ECL can be generated by triplet–triplet annihilation (T-route), which provides an alternate route to the emissive species in the singlet excited state.⁹ Another ECL mechanism involves emission from an excited state dimer such as an excimer or exciplex (E-route).^{10–13} Exciplex emission is often encountered for

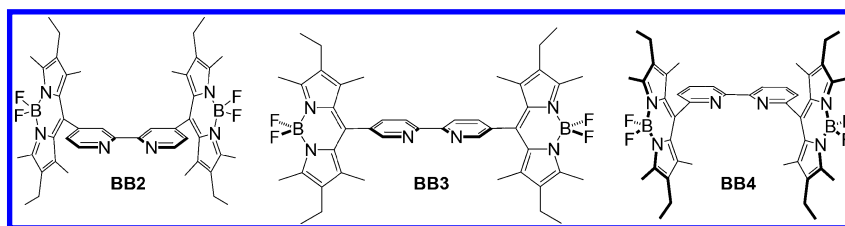
aromatic molecules, including derivatives of thianthrene,¹⁴ stilbene,¹⁵ corannulene,¹⁶ perylene,¹⁷ anthracene,^{18,19} and naphthalene²⁰ and is characterized by broad, structureless emission bands at long wavelengths.

4,4-Difluoro-4-bora-3a,4a-diaza-s-indacene (BODIPY) dyes were discovered in 1968 by Treibs and Kreuzer.²¹ BODIPY dyes are characterized by strong UV–vis absorption profiles and fluorescence quantum yields that can approach unity.^{22–24} Moreover, the photophysical properties of BODIPY fluorophores are relatively insensitive to solvent polarity and pH and they are reasonably stable under physiological conditions.^{25,26} Based on these attractive properties, BODIPY derivatives have found utility as laser dyes,²⁷ photosensitizers,^{28–30} fluorescent labels for in vivo imaging,^{31–33} and in optical devices.²⁵ The electrochemical and ECL properties of BODIPY derivatives are also of note, as these compounds often display reversible redox behavior in aprotic solvents and excellent stability upon oxidation or reduction. These attractive electrochemical properties, coupled with robust and tunable photophysics,

Received: July 2, 2013

Published: August 27, 2013

Chart 1. Structures of BODIPY-Appended Bipyridine Derivatives



distinguish BODIPY derivatives as excellent candidates for ECL applications.^{34–36}

A number of electrochemical^{37–39} and ECL studies^{40–45} of BODIPY dyes have been reported. In a recent study, we described the electrochemical, photophysical, and ECL properties of a 2,2'-bipyridine-BODIPY derivative (**BB2**), in which the meso-position of two fully substituted BODIPY moieties were linked directly to a 2,2'-bipyridine (bpy) spacer at the 4- and 4'-positions.⁴⁶ This BODIPY-appended bipyridine derivative was highly fluorescent and produced strong ECL emission in the presence of a coreactant. Given that the electronic properties of bpy derivatives are sensitive to the substitution pattern about the bpy framework, we were curious as to whether the photophysics and ECL emission might be tuned by altering the position to which the BODIPY moieties were attached. To this end, we have developed two new BODIPY-appended bipyridine derivatives, in which the BODIPY units were linked to the bpy spacer at either the 5,5'- (**BB3**) or 6,6'-positions (**BB4**). These new compounds, which are juxtaposed with our previously reported bpy-BODIPY derivative (**BB2**) in Chart 1, were thoroughly characterized by a combination of photophysical, electrochemical, and ECL methods. Of note is our finding that **BB3** and **BB4** exhibit a bifurcated ECL signal which is distinct from that simply observed by photoluminescence experiments.

EXPERIMENTAL SECTION

General Materials and Methods. Reactions were performed in oven-dried round-bottomed flasks unless otherwise noted. Reactions that required an inert atmosphere were conducted under a positive pressure of N₂ using flasks fitted with Suba-Seal rubber septa or in a nitrogen filled glovebox. Air and moisture sensitive reagents were transferred using standard syringe or cannula techniques. Reagents and solvents for synthesis were purchased from Sigma Aldrich, Acros, Fisher, Strem, or Cambridge Isotopes Laboratories. Solvents for synthesis were of reagent grade or better and were dried by passage through activated alumina and then stored over 4 Å molecular sieves prior to use.³⁹ Column chromatography was performed with 40–63 μm silica gel with the eluent reported in parentheses. Analytical thin-layer chromatography (TLC) was performed on precoated glass plates and visualized by UV or by staining with KMnO₄.

Anhydrous dichloromethane (CH₂Cl₂, ≥ 99.8%) and tetrahydrofuran (THF, ≥ 99.9%) for electrochemistry were obtained from Sigma-Aldrich (St. Louis, MO) and transferred directly into an argon atmosphere glovebox (MBraun Inc., Stratham, NH) without further purification. Electrochemical grade tetra-*n*-butylammonium hexafluorophosphate (TBAPF₆) was dried in a vacuum oven at 100 °C prior to transferring directly into an argon atmosphere glovebox. Benzoyl peroxide (BPO) and tri-*n*-propylamine (TPA) were obtained from Sigma-Aldrich and were used as received.

Compound Characterization. ¹H and ¹³C NMR spectra were recorded at 25 °C on a Bruker 400 MHz spectrometer. Proton spectra are referenced to the residual proton resonance of the deuterated solvent (CDCl₃ = δ 7.26; DMSO-*d*₆ = δ 2.50), and carbon spectra are referenced to the carbon resonances of the solvent (CDCl₃ = δ 77.23; DMSO-*d*₆ = δ 40.45). All chemical shifts are reported using the

standard δ notation in parts-per-million; positive chemical shifts are to a higher frequency from the given reference. LR-GCMS data were obtained using an Agilent gas chromatograph consisting of a 6850 Series GC System equipped with a 5973 Network Mass Selective Detector. Low resolution MS data were obtained using either an LCQ Advantage from ThermoFinnigan or a Shimadzu LC/MS-2020 single quadrupole MS coupled with an HPLC system, with a dual ESI/APCI source. High-resolution mass spectrometry analyses were either performed by the Mass Spectrometry Laboratory in the Department of Chemistry and Biochemistry at the University of Delaware or at the University of Illinois at Urbana–Champaign.

2,2'-Bipyridine-5,5'-dicarboxylic Acid (1). This compound was prepared using a slightly modified literature method.⁴⁷ 5,5'-Dimethyl-2,2'-bipyridine (4.69 g, 0.025 mol) and potassium permanganate (26.2 g, 0.17 mol) were added to 300 mL of water. The resulting mixture was heated at 90 °C with stirring under air for 14 h. After the mixture cooled to room temperature, a brown precipitate was removed by filtration and the filtrate was extracted three times with Et₂O. The resulting aqueous solution was acidified to a pH of 2 with 1 M HCl, which led to formation of a white precipitate, which was collected by vacuum filtration to deliver the title compound (4.27 g, 70%). ¹H NMR (400 MHz, DMSO, 25 °C) δ/ppm: 9.22 (s, 2H), 8.60 (d, *J* = 8.3 Hz, 2H), 8.49 (s, 2H). HR-ESI-MS [M+H]⁺ *m/z*: calcd for C₁₂H₉N₂O₄, 245.0562. Found 245.0558.

5,5'-Bis(BODIPY)-2,2'-bipyridine (BB3). This compound was prepared by adapting a literature method.⁴⁶ 2,2'-Bipyridine-4,4'-dicarboxylic acid (**1**) (0.5 g, 2.23 mmol) was suspended in 20 mL of SOCl₂. The resulting mixture was heated to reflux under nitrogen with stirring for 24 h, after which time all solid materials had dissolved to produce a dark yellow solution. The SOCl₂ was removed under reduced pressure, and the resulting residue was dissolved in 100 mL of CHCl₃ and sparged with nitrogen for 45 min. 2,4-Dimethyl-3-ethylpyrrole (2.11 mL, 15.6 mmol) was added to the solution, and the reaction was heated at 50 °C under nitrogen for 90 min. After the solution cooled to room temperature, the solvent was removed by rotary evaporation and the resulting residue was dissolved in 200 mL of toluene and CH₂Cl₂ (95:5). Following the addition of triethylamine (2.48 mL, 17.8 mmol), the resulting dark colored solution was stirred under air for 30 min at room temperature. The solution was then heated to 50 °C, and 3.85 mL (31.2 mmol) of BF₃·OEt₂ were added. The reaction was stirred for an additional 90 min at 50 °C, after which time the solvent was removed under reduced pressure. The resulting residue was purified via flash column chromatography on silica using CH₂Cl₂ and MeOH (99:1) as the eluent to deliver 746 mg (44%) of the title compound as a brick red powder. ¹H NMR (400 MHz, CDCl₃, 25 °C) δ/ppm: 8.66 (s, 2H), 8.64 (d, *J* = 4.0 Hz 2H), 7.83 (d, 2H), 2.56 (s, 12H), 2.32 (q, 8H), 1.40 (s, 12H), 1.00 (t, 12H). ¹³C NMR HR-ESI-MS [M+H]⁺ *m/z*: calcd for C₄₄H₅₂B₂F₄N₆, 761.4297. Found 761.4297.

2,2'-Bipyridine-6,6'-dicarboxylic Acid (2). This compound was prepared using the same method used for the preparation of 2,2'-bipyridine-5,5'-dicarboxylic acid (**1**), using 6,6'-dimethyl-2,2'-bipyridine in place of 5,5'-dimethyl-2,2'-bipyridine. The desired product was isolated in 70% yield. ¹H NMR (400 MHz, DMSO, 25 °C) δ/ppm: 8.78 (d, *J* = 7.7 Hz, 2H), 8.22 (t, *J* = 8.0 Hz, 2H), 8.17 (d, *J* = 8.0 Hz, 4H). ¹³C NMR (101 MHz, DMSO, 25 °C) δ/ppm: 165.92, 154.46, 148.10, 138.94, 125.24, 124.10. HR-ESI-MS [M+H]⁺ *m/z*: calcd for C₁₂H₉N₂O₄, 245.0562. Found 245.0565.

6,6'-Bis(BODIPY)-2,2'-bipyridine (BB4). This compound was prepared according to the same method used for the preparation of 5,5'-Bis(BODIPY)-2,2'-bipyridine (BB3), using 2,2'-bipyridine-6,6'-dicarboxylic acid (2) in place of 2,2'-bipyridine-5,5'-dicarboxylic acid (1). The desired product was purified via flash column chromatography on silica using CH_2Cl_2 as the eluent to deliver 610 mg (36%) of the title compound as a brick red powder. $^1\text{H NMR}$ (400 MHz, CDCl_3 , 25 °C) δ /ppm: 8.49 (d, $J = 8.0$ Hz, 2H), 7.92 (t, $J = 7.8$ Hz, 2H), 7.45 (d, $J = 6.8$ Hz, 2H), 2.56 (s, 12H), 2.31 (q, $J = 7.5$ Hz, 8H), 1.27 (s, 13H), 0.99 (t, $J = 7.5$ Hz, 12H). $^{13}\text{C NMR}$ (101 MHz, CDCl_3 , 25 °C) δ 156.05, 154.67, 154.16, 138.04, 137.77, 137.26, 132.96, 130.74, 124.82, 121.43, 17.09, 14.68, 12.68, 11.49. HR-ESI-MS $[\text{M}+\text{H}]^+$ m/z : calcd for $\text{C}_{44}\text{H}_{52}\text{B}_2\text{F}_4\text{N}_6$, 761.4297. Found 761.4301.

Electrochemical Experiments. Electrochemistry experiments were carried out using a three-electrode setup that employed a 0.043 cm^2 platinum disk working electrode (WE), a platinum counter electrode (CE), and a silver wire quasi-reference electrode (RE) calibrated using ferrocene as an internal standard (0.342 V vs SCE).⁴⁸ An L-shaped electrode (disk oriented vertically) was used for the CV measurements and ECL experiments. The working electrode was polished prior to every experiment with $0.3\ \mu\text{m}$ alumina particles dispersed in water, followed by sonication in ethanol and water for several minutes. All glassware was oven-dried for 1 h at 120 °C prior to transferring into an argon atmosphere glovebox. All solutions were prepared inside the glovebox and sealed in a vacuum-tight electrochemical cell with a Teflon screw cap containing three metal rods for electrode connections. Cyclic voltammetry and chronoamperometry experiments were carried out with a CH Instruments (Austin, TX) model 660 electrochemical workstation. Electrochemistry experiments were carried out in CH_2Cl_2 containing 0.1 M TBAPF₆ as the supporting electrolyte.

ECL Experiments. ECL spectra were generated by annihilation by pulsing the potential with a pulse width of 0.1 s from about 80 mV past the peak potentials, or by stepping from 0 to 80 mV from the reduction peak with a step time of 2 s using benzoyl peroxide as a coreactant or by stepping from 0 V to 80 mV from the oxidation peak with a step time of 2 s using TPA as a coreactant. ECL spectra were recorded with a Princeton Instruments Spec 10 CCD camera (Trenton, NJ) with an Acton SpectPro-150 monochromator cooled with liquid nitrogen to -100 °C .

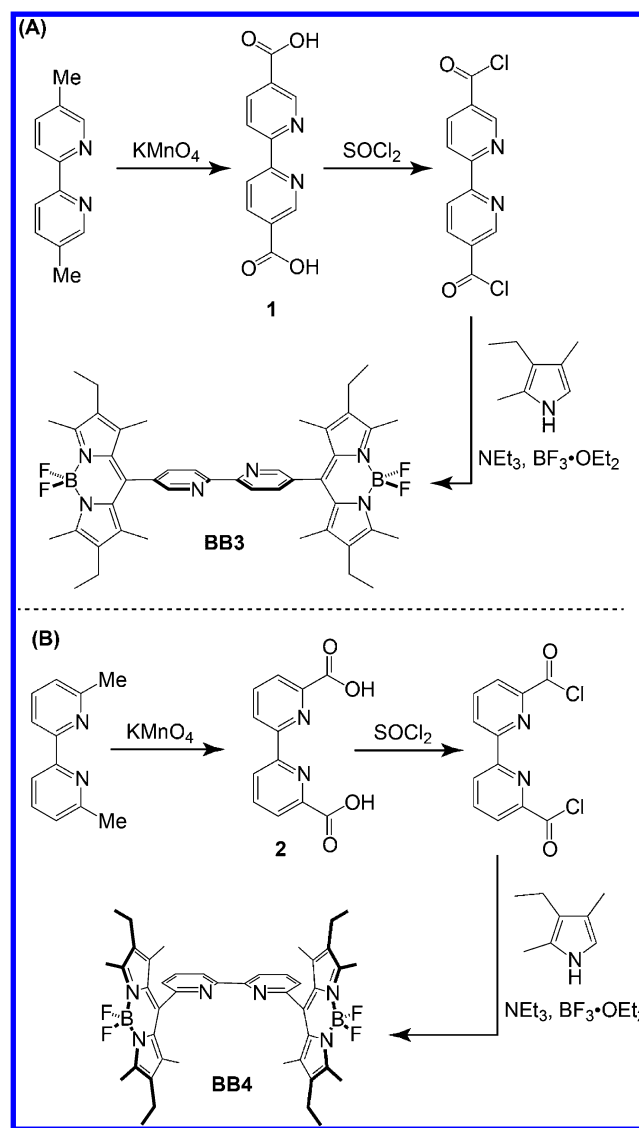
ECL-CV simultaneous experiments were carried out prior to spectral measurements to ensure the presence of ECL emission. In this case, a multichannel Eco Chemie Autolab PGSTAT100 (Utrecht, The Netherlands) was used to collect the signal and a photomultiplier tube (Hamamatsu R4220, Tokyo, Japan) was used as a detector. Voltage for the PMT (750 V) was provided by a Kepco power supply (New York, NY), and the signal from the PMT to the potentiostat was transferred using a multimeter (Keithley, Solon, OH). Digital simulations were performed using Digisim computer software (Bioanalytical Systems, West Lafayette, IN).

Computations. Geometry optimizations, frequency calculations, and molecular orbital calculations were performed in Gaussian 09 using the B3LYP/6-311G(d) basis set. Only positive frequencies were found for the optimized structures. Molecular orbitals were visualized using Gaussview software. All calculations were performed in the gas phase.

RESULTS AND DISCUSSION

Synthesis. The synthetic protocols used to construct the bpy-BODIPY derivatives 5,5'-bis(BODIPY)-2,2'-bipyridine (BB3) and 6,6'-bis(BODIPY)-2,2'-bipyridine (BB4) are shown in Scheme 1. Preparation of BB3 begins with the conversion of 5,5'-dimethyl-2,2'-bipyridine to the corresponding dicarboxylic acid (1) via oxidation with KMnO_4 in water (Scheme 1a). Following purification, the dicarboxylic acid derivative was treated with excess thionyl chloride to generate the corresponding acid chloride derivative. Condensation of this intermediate with 2,4-dimethyl-3-ethylpyrrole followed by

Scheme 1. Synthesis of BODIPY-Appended Bipyridine Derivatives BB3 and BB4



treatment with NEt_3 and $\text{BF}_3 \cdot \text{OEt}_2$ afforded 5,5'-bis(BODIPY)-2,2'-bipyridine (BB3) in 44% yield. Synthesis of the homologous 6,6'-bis(BODIPY)-2,2'-bipyridine (BB4) derivative was accomplished in 36% yield from dicarboxylic acid 2 using an analogous strategy (Scheme 1b).

Electrochemistry. The basic redox properties of BB3 and BB4 were probed by cyclic voltammetry (CV). Electrochemical results obtained for both bpy-BODIPY derivatives in CH_2Cl_2 containing 0.1 M TBAPF₆ as the supporting electrolyte are summarized in Table 1, and pertinent CVs are shown in Figure 1. As was previously observed for BB2, both BB3 and BB4 show single redox waves upon oxidation or reduction. Both these waves consist of two closely spaced one-electron ($1e^-$) events, as confirmed by digital simulation. Overall $2e^-$ reduction and oxidation waves were also observed for BB3 and BB4 using a platinum ultramicroelectrode (Figure S1). Fitting of the CV traces was carried out by subtraction of the background current and determining the best fit to the simulation corrected for the measured electrical double layer capacitance and the uncompensated cell resistance (Supporting Information Figures S4–S7).

Table 1. Redox Potentials Recorded for BB2–BB4 in CH₂Cl₂^a

	$E_{1/2}$ (A/A ⁺)	$E_{1/2}$ (A ⁺ /A ²⁺)	$E_{1/2}$ (A/A ⁻)	$E_{1/2}$ (A ⁻ /A ²⁻)	$D \times 10^6$	ΔG_{ann}^c	ΔH_{ann}^d
BB2 ^b	1.11 V	1.15 V	-1.15 V	-1.22 V	4.0 cm ² /s	2.33 eV	2.23 eV
BB3	1.14 V	1.19 V	-1.17 V	-1.24 V	4.0 cm ² /s	2.31 eV	2.21 eV
BB4	1.12 V	1.17 V	-1.18 V	-1.24 V	4.0 cm ² /s	2.30 eV	2.20 eV

^a $E_{1/2}$ values were obtained by fitting the experimental results to digital simulations and are reported versus SCE. ^bData for BB2 are reproduced from ref 46. ^c $\Delta G_{\text{ann}} = E_{1/2}(\text{A/A}^+) - E_{1/2}(\text{A/A}^-)$. ^d $\Delta H_{\text{ann}} = \Delta G_{\text{ann}} - 0.1$ eV.

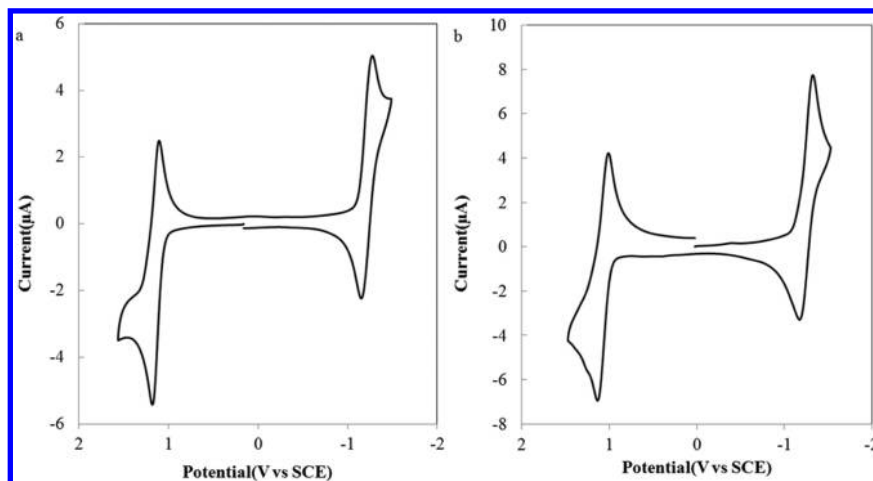


Figure 1. Cyclic voltammograms of CH₂Cl₂ solutions of (a) 0.59 mM BB3 and (b) 0.68 mM BB4. CVs were recorded using 0.1 M TBAPF₆ as the supporting electrolyte using a platinum disk working electrode ($A = 0.043$ cm²) and a scan rate (ν) of 100 mV/s.

The small separation observed for the two reductive and oxidative electron transfer events is consistent with there being negligible electronic coupling between the two BODIPY moieties via the bpy spacers of both BB3 and BB4. For two groups that are completely uncoupled, the theoretical difference between the potentials for the first and second electron transfers would be 36 mV due to entropic factors.⁴⁹ As was the case for BB2, the weak electronic coupling between the two BODIPY groups of BB3 and BB4 is manifest in small separations in potential between the A/A⁺ and A⁺/A²⁺ redox waves of approximately 50–70 mV, as judged by the digital simulations described above (Table 1). If the BODIPY groups were more strongly coupled, the two oxidation and reduction waves would be more broadly separated with the A/A²⁺ and A⁻/A²⁻ shifted toward more extreme potentials,⁵⁰ as has been observed for BODIPY derivatives containing directly linked indacene moieties.³⁴

DFT calculations carried out for BB3 and BB4 confirm that the BODIPY moieties of both these molecules are effectively insulated from one another, as steric demands force the indacene frameworks to cant significantly with respect to the bipyridine bridges. Orthogonalization of the BODIPY moieties and bpy spacer results in the HOMO and LUMO of both BB3 and BB4 residing on the individual chromophores. These calculations provide no evidence for electronic delocalization onto the bipyridine spacer (Figure 2).

Oxidation and reduction of both BB3 and BB4 are reversible, which is consistent with the high stability of the radical cation and anion products formed at the electrode surface. This stability is a result of the BODIPY units of both bpy-BODIPY homologues being completely substituted along the indacene periphery, which prevents decomposition of the electrochemically generated radical ions. Randles–Sevcik analysis showed that the measured oxidation and reduction peaks vary linearly

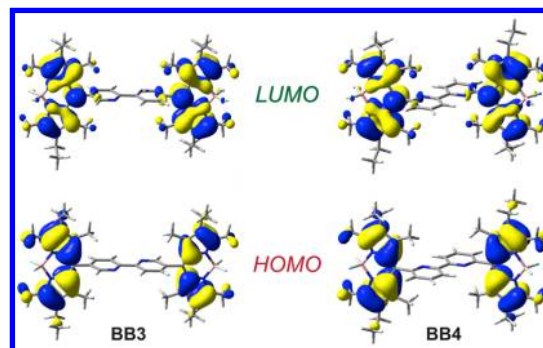


Figure 2. Calculated frontier molecular orbitals for BB3 and BB4 by DFT (B3LYP/6-311G(d)).

as a function of the square root of the scan rate ($\nu^{1/2}$) (Supporting Information Figures S2 and S3), indicating that these redox processes are diffusion controlled.⁵¹ In an effort to obtain better insight into the mechanism and kinetics of both the anodic and cathodic processes, the experimental polarization curves for BB3 and BB4 were digitally simulated (Supporting Information Figures 3 and S4–S7). Both the cathodic and anodic CVs of BB3 were characterized by two reversible 1e⁻ transfers, with a fast heterogeneous rate constant ($k^0 > 0.01$ cm/s) for both reaction steps (Supporting Information Figures 3a, b, S4, and S5). Virtually identical parameters were also found for BB4 (Supporting Information Figures 3c, d, S6, and S7). The slight differences between simulated and experimental polarization curves may be attributed to weak adsorption of the bpy-BODIPY derivatives to the working electrode. BODIPY redox sites can often be oxidized or reduced by two electron equivalents. As such, the bpy-BODIPY constructs considered here might be expected to be able to donate up to four electrons at sufficiently positive

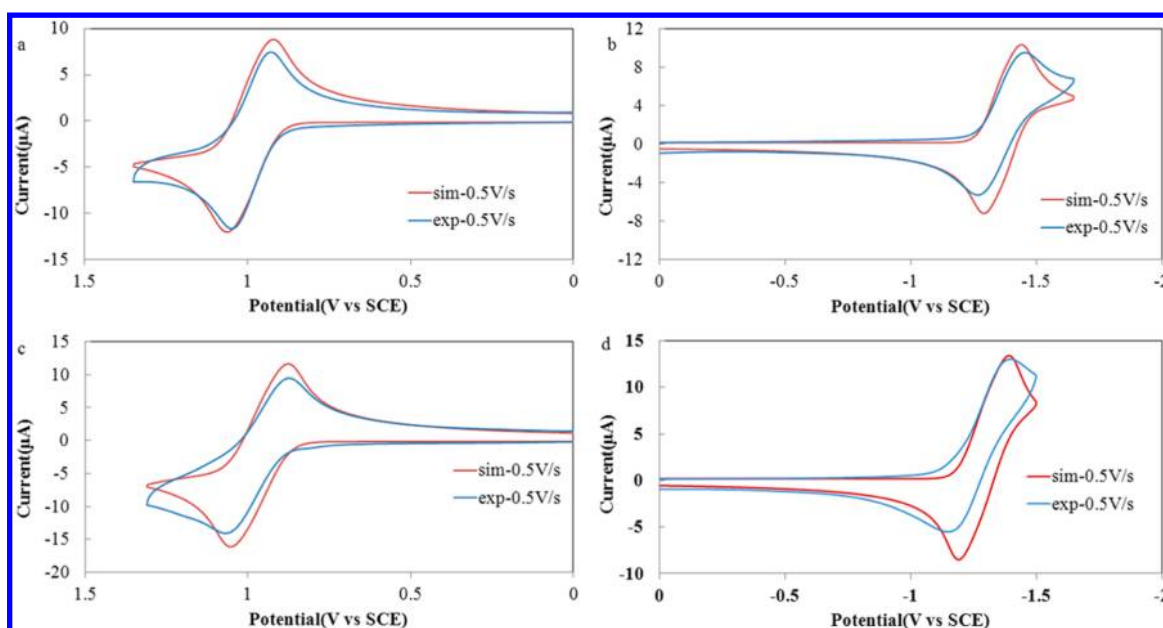


Figure 3. Comparison between simulated (red) and experimental (blue) polarization curves for (a, b) **BB3** and (c, d) **BB4**. Simulations were prepared for an EE mechanism with a heterogeneous ET rate constant of $k^{\circ} > 0.01$ cm/s. Experimental polarization curves recorded in CH_2Cl_2 containing 0.1 M TBAPF₆ with a platinum disk working electrode ($A = 0.043$ cm²) at a scan rate (ν) of 500 mV/s.

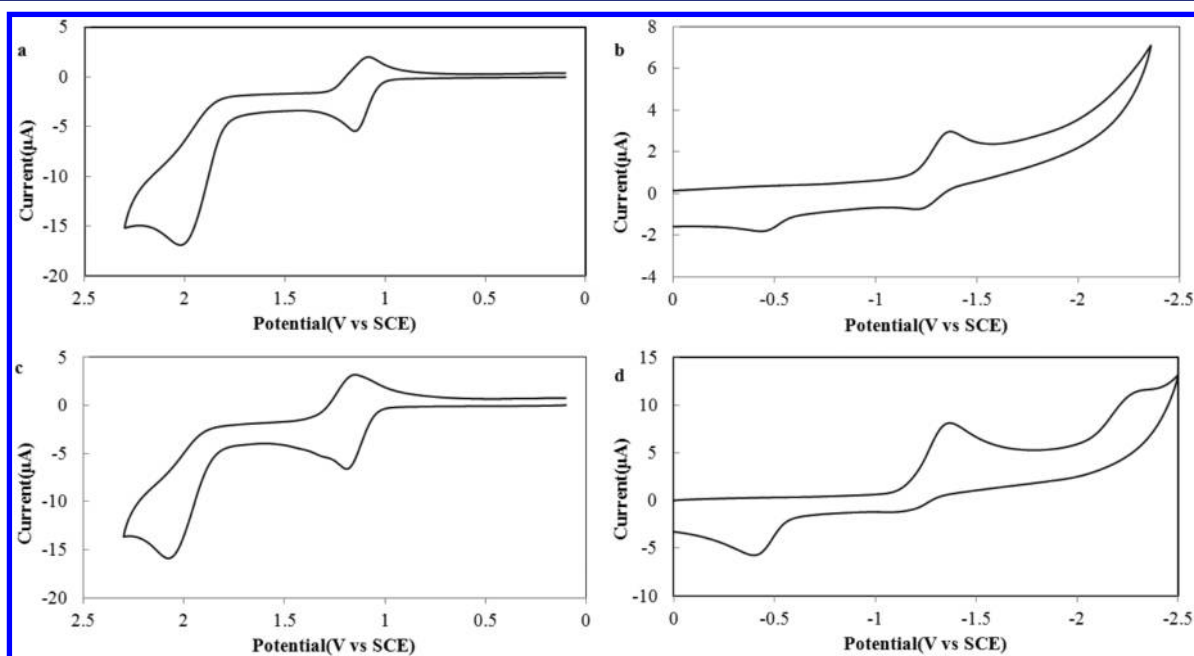


Figure 4. Cyclic voltammograms of (a, b) **BB3**, and (c, d) of **BB4**. Anodic scans of (a) 0.59 mM **BB3** and (c) 0.68 mM **BB4** were recorded in CH_2Cl_2 containing 0.1 M TBAPF₆. Cathodic scans of (b) 0.56 mM **BB3** and (d) 0.78 mM **BB4** were recorded in THF containing 0.1 M TBAPF₆. All polarization curves were recorded using a platinum disk working electrode ($A = 0.043$ cm²) at a scan rate (ν) of 100 mV/s.

potentials. CV scans recorded for **BB3** and **BB4** in CH_2Cl_2 over the potential range of 0.1–2.4 V versus SCE revealed a second irreversible peak at ~ 2.05 V (Figure 4). This second oxidation wave is more positive than the first $2e^-$ wave by ~ 0.9 V, which is consistent with that observed for other BODIPY derivatives^{28,46} and is presumed to correspond to removal of a third and fourth e^- from the bpy-BODIPY derivatives. The reductive window of CH_2Cl_2 precludes scanning to potentials significantly more negative than the first reduction waves; however, THF, which has a more negative working potential range, was used to study the reduction of **BB3** and **BB4** beyond

–1.5 V. An additional reduction wave was observed for **BB4** at approximately –2.3 V versus SCE; however, **BB3** did not show a new redox wave beyond the first $2e^-$ reduction. This divergent behavior is likely a result of the distinct molecular topologies of the two bpy-BODIPY homologues (Chart 1).

Photophysics. The basic photophysical properties of **BB3** and **BB4** were assessed in CH_2Cl_2 (Figure 5) and are summarized in Table 2. Both **BB3** and **BB4** display optical properties analogous to those of the previously studied **BB2** homologue and other related BODIPY derivatives. UV–vis absorbance and emission spectra for **BB3** and **BB4** are shown

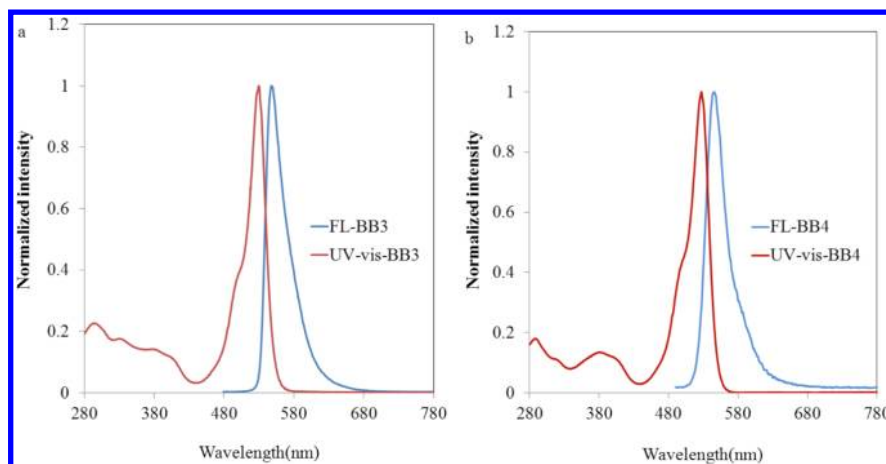


Figure 5. Normalized absorbance (red) and emission (blue) spectra for (a) **BB3** and (b) **BB4** in CH_2Cl_2 . Emission samples were excited at $\lambda_{\text{ex}} = 480$ nm.

Table 2. Photophysical and ECL Properties Recorded for BB2–BB4 in CH_2Cl_2

	absorbance		emission			electrogenerated chemiluminescence	
	λ_{abs}	ϵ ($\text{M}^{-1} \text{cm}^{-1}$)	λ_{Fl}	Φ_{Fl}	E_{0-0}^a	λ_{ECL}	Φ_{ECL}^c
BB2 ^b	528 nm	8.4×10^4	547 nm	0.39	2.26 eV	554 nm	–
BB3	528 nm	8.2×10^4	545 nm	0.39	2.27 eV	570 nm, 740 nm	0.006
BB4	528 nm	6.7×10^4	543 nm	0.47	2.28 eV	571 nm, 741 nm	0.010

^a E_{0-0} approximate energy of the first singlet excited state taken as the fluorescence wavelength maximum. ^bData for **BB2** are reproduced from ref 46. ^cECL efficiencies were determined using $[\text{Ru}(\text{bpy})_3]^{2+}$ in MeCN as a standard, for which $\Phi_{\text{ECL}} = 0.05$.

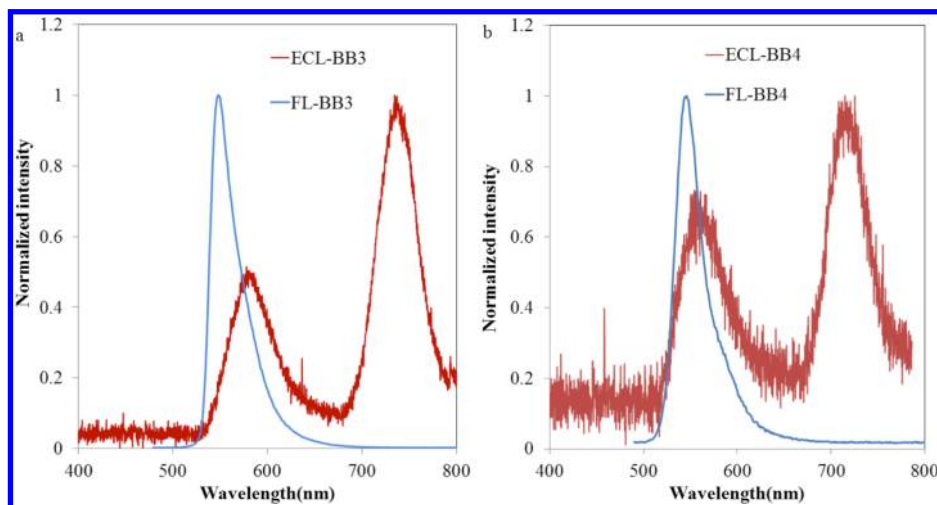


Figure 6. Fluorescence (blue) and ECL (red) spectra of (a) **BB3** and (b) **BB4**. ECL emission was generated by oscillating the potential with a pulse width of 0.1 s from ~ 80 mV past the first oxidation and reduction waves of each Bpy-BODIPY homologue. ECL spectra were recorded in CH_2Cl_2 containing 0.1 M TBAPF₆ with an accumulation time of 2 min.

in Figure 5. Both of these compounds display strong absorption bands in the visible region centered at approximately 528 nm ($S_0 \rightarrow S_1$ transition) and 371 nm ($S_0 \rightarrow S_2$ transition).²⁵ The molar absorption coefficients of **BB3** and **BB4** in CH_2Cl_2 were measured to be 82 000 and 67 000 $\text{M}^{-1} \text{cm}^{-1}$, respectively. These relatively high absorptivities are consistent with there being two BODIPY chromophores appended to the bipyridine bridge for both compounds. These molar absorptivities are similar in magnitude to that determined for **BB2** (Table 2).⁴⁶ The bipyridine spacer of **BB3**–**BB4** causes the major absorbance band observed for each of these derivatives to undergo a hypsochromic shift of approximately 7 nm relative to

the analogous BODIPY dye bearing a methyl substituent at the indacene meso-position (PM567).⁵² These hypsochromic shifts can be attributed to an inductive electron withdrawing effect of the bipyridine spacer.⁵³

Excitation of **BB3** and **BB4** leads to a greenish-yellow emission, centered at 545 and 543 nm, respectively (Figure 5, Table 2). The small Stokes shift observed for both bpy-BODIPY derivatives was also observed for **BB2**⁴⁶ and is typical of many BODIPY dyes.⁵⁴ Measured quantum yields of fluorescence (Φ_{Fl}) for **BB3** and **BB4** (0.39 and 0.47, respectively) are also similar to that determined for **BB2**.⁴⁶ Notably, there was no evidence for dimerization or aggregation

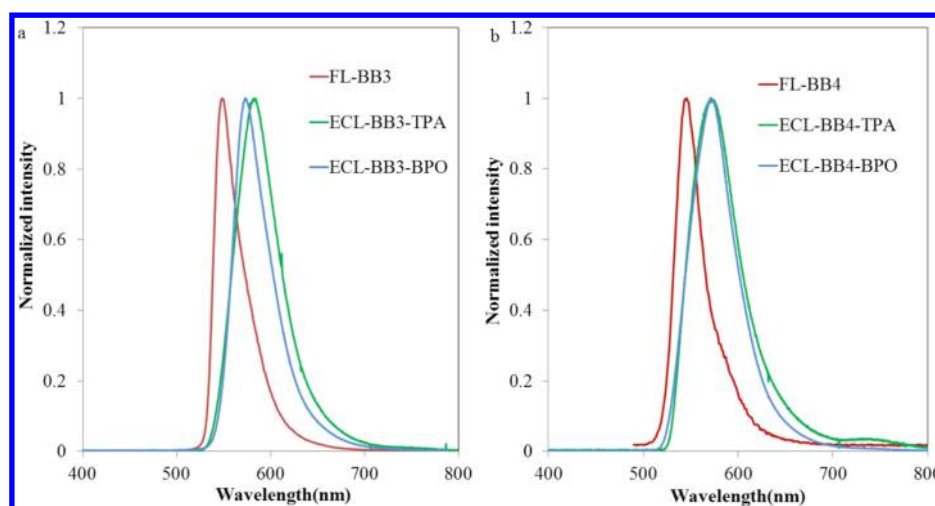
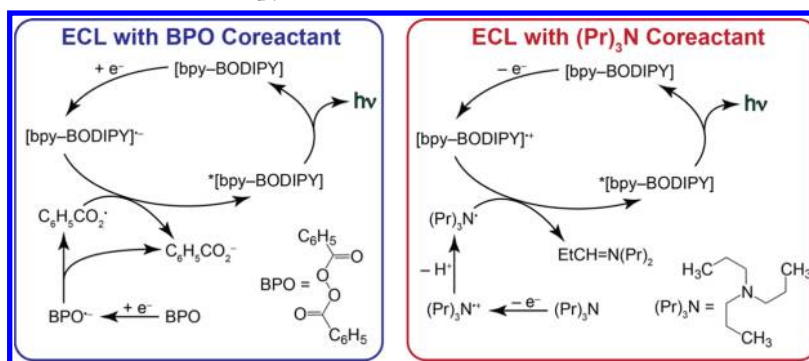


Figure 7. Normalized fluorescence (red) and ECL (blue and green) spectra recorded for 0.5 mM solutions of (a) BB3 and (b) BB4 in CH_2Cl_2 containing 0.1 M TBAPF₆. ECL spectra in blue were generated by pulsing the applied potential from 0 to -1.3 V versus Ag/Ag^+ in the presence of 3.5 mM benzoyl peroxide. ECL spectra in green were generated by pulsing the applied potential from 0 to 1.3 V versus Ag/Ag^+ in the presence of 25 mM TPA.

Scheme 2. Pathways for ECL Generation from bpy-BODIPY Derivatives in the Presence of Either BPO or $(\text{Pr})_3\text{N}$ Coreactants



of the bpy-BODIPY complexes in solution, as the shape and normalized intensity of the photoluminescence spectra recorded for BB3 and BB4 did not vary as a function of dye concentration.

Electrogenerated Chemiluminescence. Initial ECL studies for BB3 and BB4 were carried out by pulsing between the first reduction and oxidation potentials (80 mV past both the reduction and oxidation waves). ECL spectra obtained via annihilation of the ions and diions generated during these experiments are shown in Figure 6, and associated spectral data are listed in Table 2. Two ECL emission peaks are observed for both bpy-BODIPY homologues. Higher-energy ECL bands were observed at ~ 570 nm, which are similar in energy to those observed in the photoluminescence spectrum. Lower-energy ECL signals were also resolved at ~ 740 nm. By contrast, when the excited states of BB3 or BB4 were generated under oxidizing conditions using tripropylamine (TPA) as a coreactant, or under reducing conditions using benzoyl peroxide (BPO) as a coreactant, only the high-energy ECL peaks ($\lambda_{\text{ECL}} \approx 570$ nm) were observed (Figure 7). These ECL emission profiles are very similar to the normal fluorescence spectra observed for BB3 and BB4 when corrected for a small difference inner filter effect (Figure 7).⁴⁶ Virtually identical results were obtained previously for BB2, suggesting that all three bpy-BODIPY derivatives produce ECL via the common pathways shown in Scheme 2.⁴⁶

In order to clarify the nature of the excited state from which ECL emission is produced for BB3 and BB4, the enthalpy of annihilation (ΔH_{ann}) was assessed for both compounds. This parameter was estimated from the reversible standard potentials for the reduction and oxidation of each of the bpy-BODIPY homologues according to the expression $\Delta H_{\text{ann}} = \Delta G_{\text{ann}} - T\Delta S$. The free energy of annihilation (ΔG_{ann}) was calculated as the difference between $E_{1/2}(\text{A}/\text{A}^+)$ and $E_{1/2}(\text{A}/\text{A}^-)$, and $T\Delta S$ was estimated to be 0.1 eV,⁵⁵ leading to an estimation of $\Delta H_{\text{ann}} \approx 2.20$ eV. The energies of the first singlet excited state (E_{0-0}) of BB3 and BB4 were also determined based on their maximal fluorescence wavelength (λ_{Fl}) to be $E_{0-0} = 2.27$ eV. The estimated values of ΔH_{ann} and E_{0-0} for BB3 and BB4 differ by less than 0.1 eV, which suggests that the bpy-BODIPY ECL and fluorescence originate from the same excited state. Moreover, these results suggest that the ECL signal observed at ~ 570 nm for BB3 and BB4 is generated through either an S- or a T-route (vide supra).

In addition to the ECL signal at ~ 570 nm, long-wavelength emission at ~ 740 nm was also observed under annihilation conditions. This long-wavelength signal was not observed in the photoluminescence spectra recorded for BB3 and BB4. As has been observed for previously studied BODIPY derivatives,³⁴ long-wavelength ECL can be generated via formation of excimers; however, such signals are usually broad and structureless. That the ECL signal observed for BB3 and

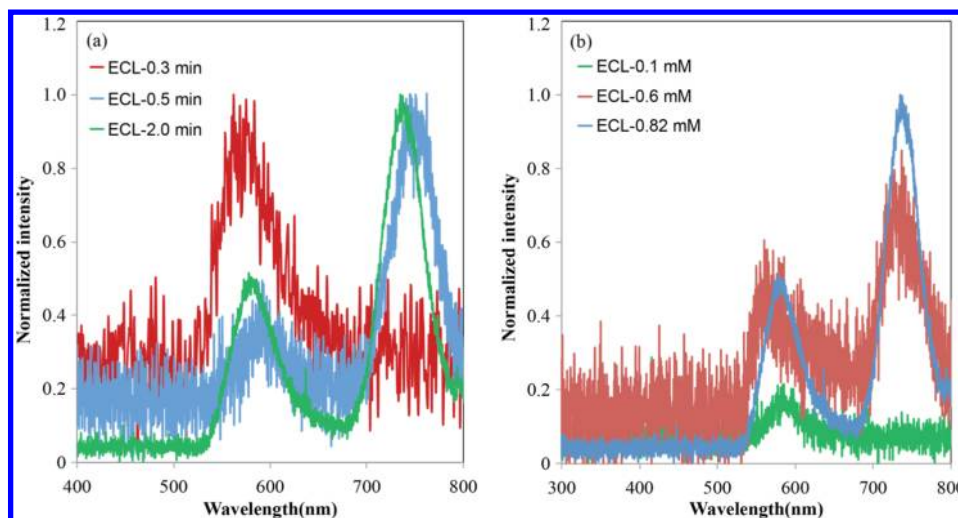


Figure 8. Annihilation ECL spectra recorded (a) as a function of accumulation time for 0.82 mM **BB3** and (b) with varying concentrations of **BB3**.

BB4 at 740 nm is relatively sharp suggests that this emission is generated via an alternate pathway.

A second pathway by which long-wavelength ECL can be obtained involves electrochemical production of side products, which in and of themselves are efficient ECL luminophores in the red region. Such phenomena have been observed in other ECL studies involving BODIPY derivatives.^{40,56} In order to determine if a bpy-BODIPY decomposition product was responsible for the long-wavelength ECL in Figure 6, we examined the time and concentration dependence for the ECL signal at 740 nm. As shown in Figure 8a, the ratio of the ECL signal produced by **BB3** at 740 nm versus that at 570 nm (I_{740}/I_{570}) increases with ECL accumulation time. Additionally, when the annihilation ECL experiment was repeated with **BB3** solutions of varying concentration (0.1–0.8 mM), the signal at 740 nm was maximized at higher concentrations (Figure 8b). When taken together, these experiments suggest that the ECL band at 740 nm is generated by a new species formed from **BB3** during the annihilation process. We note that formation of this new species requires the annihilation reaction and is not simply the result of decomposition of one of the radical ions formed during the experiment. Moreover, the rise in ECL intensity at 740 nm is concomitant with a change in the color of the ECL solution from orange to deep purple. This change is irreversible, as the color of the solution does not revert back to orange after completion of the ECL experiment. These observations suggest that the long-wavelength ECL signal is generated from a new emissive product that is irreversibly generated from **BB3** during the annihilation experiment. Future investigations will focus on elucidation of these electrochemical products, which may be composed of bpy-BODIPY dimers or oligomers.

SUMMARY

Two new 2,2'-bipyridine derivatives containing ancillary BODIPY units at either the 5- and 5'- (**BB3**) or 6- and 6'- positions (**BB4**) were prepared, and the basic photophysics, electrochemistry, and ECL properties of these bpy-BODIPY homologues were investigated. These properties are similar to those observed for a related bpy-BODIPY derivative in which two BODIPY moieties were coupled to a 2,2'-bipyridine spacer at the 4- and 4'-positions (**BB2**). All three of the bpy-BODIPY constructs are strongly absorbing in the visible region and

display high fluorescence quantum yields in CH_2Cl_2 . Cyclic voltammetry showed that the new bpy-BODIPY derivatives (**BB3** and **BB4**) also maintain redox properties that are similar to that observed for **BB2**. All three systems display reversible $2e^-$ oxidation and reduction waves, which consist of two closely spaced (50–70 mV) $1e^-$ events. These observations are consistent with the calculated frontier molecular orbitals for these compounds and indicate that the 2,2'-bipyridine spacer of each bpy-BODIPY homologue does not facilitate efficient electronic communication between the tethered indacene units, regardless of their position on the bpy bridge.

ECL experiments for **BB3** and **BB4** correlate with the observed electrochemistry. In the presence of a coreactant (TPA or BPO) both of these bpy-BODIPY derivatives exhibit strong ECL and produce spectra that are very similar to their corresponding fluorescence profiles. Similar results have been observed for **BB2**. Under annihilation conditions, however, the ECL signal obtained from **BB3** and **BB4** is distinct and is characterized by two major bands. One of these bands is centered at ~ 570 nm and is attributed to emission via an S- or a T-route. The second band occurs at longer wavelengths and is centered around ~ 740 nm. This long-wavelength ECL signal was not observed in the **BB3** or **BB4** photoluminescence spectra. The shape and concentration dependence of this long-wavelength ECL band was not indicative of luminescence from an excimer or aggregate, but rather is generated by a new species formed from the bpy-BODIPY luminophores during the annihilation process. Future work will be needed to elucidate the structure of these electrochemical products, which may be comprised of bpy-BODIPY dimers, oligomers, or more extended assemblies.

ASSOCIATED CONTENT

Supporting Information

Electrochemistry and spectroscopic data. This material is available free of charge via the Internet at <http://pubs.acs.org>.

AUTHOR INFORMATION

Corresponding Author

ajbard@mail.utexas.edu; joelr@udel.edu

Present Address

[§]Key Laboratory of Analytical Chemistry for Life Science of Shaanxi Province, School of Chemistry and Materials Science, Shaanxi Normal University, Xi'an, 710062, P. R. China.

Notes

The authors declare no competing financial interest.

ACKNOWLEDGMENTS

A.J.B. acknowledges support from the Robert A. Welch Foundation (F-0021) and the National Science Foundation (CHE-1111518). J.R. was supported through a DuPont Young Professor award. J.R. also thanks the University of Delaware and the Donors of the American Chemical Society's Petroleum Research Fund for financial support. H.L.Q. thanks The National Science Foundation of China (Nos. 21375084, 21027007), the Natural Science Basic Research Plan in Shaanxi Province of China (No. 2013KJXX-73), and the Fundamental Research Funds for the Central Universities (No. GK261001185) for support. NMR and other data were acquired using instrumentation obtained with assistance from the NSF (CHE-0421224, CHE-0840401, CHE-1048367 and CHE-1229234).

REFERENCES

- (1) *Electrogenerated Chemiluminescence*; Bard, A. J., Ed.; Dekker: New York, 2004.
- (2) Marcus, R. A. *J. Chem. Phys.* **1965**, *43*, 2654–2657.
- (3) Knight, A. W.; Greenway, G. M. *Analyst* **1994**, *119*, 879–890.
- (4) Armstrong, N. R.; Wightman, R. M.; Gross, E. M. *Annu. Rev. Phys. Chem.* **2001**, *52*, 391–422.
- (5) Velasco, J. G. *Electroanalysis* **1991**, *3*, 261–271.
- (6) Forster, R. J.; Bertoncello, P.; Keyes, T. E. *Annu. Rev. Anal. Chem.* **2009**, *2*, 359–385.
- (7) Richter, M. M. *Chem. Rev.* **2004**, *104*, 3003–3036.
- (8) Miao, W. J. *Chem. Rev.* **2008**, *108*, 2506–2553.
- (9) Omer, K. M.; Ku, S. Y.; Wong, K. T.; Bard, A. J. *Angew. Chem., Int. Ed.* **2009**, *48*, 9300–9303.
- (10) Maloy, J. T.; Bard, A. J. *J. Am. Chem. Soc.* **1971**, *93*, 5968–5981.
- (11) Keszthelyi, C. P.; Bard, A. J. *Chem. Phys. Lett.* **1974**, *24*, 300–304.
- (12) Park, S. M.; Bard, A. J. *J. Am. Chem. Soc.* **1975**, *97*, 2978–2985.
- (13) Chandros, E. A.; Longwort, J. W.; Visco, R. E. *J. Am. Chem. Soc.* **1965**, *87*, 3259–3260.
- (14) Keszthelyi, C. P.; Tachikawa, H.; Bard, A. J. *J. Am. Chem. Soc.* **1972**, *94*, 1522–1527.
- (15) Keszthel, C. P.; Bard, A. J. *Chem. Phys. Lett.* **1974**, *24*, 300–304.
- (16) Valenti, G.; Bruno, C.; Rapino, S.; Fiorani, A.; Jackson, E. A.; Scott, L. T.; Paolucci, F.; Marcaccio, M. *J. Phys. Chem. C* **2010**, *114*, 19467–19472.
- (17) Oyama, M.; Mitani, M.; Okazaki, S. *Electrochem. Commun.* **2000**, *2*, 363–366.
- (18) Tachikaw, H.; Bard, A. J. *Chem. Phys. Lett.* **1974**, *26*, 568–573.
- (19) Itaya, K.; Tushima, S. *Chem. Phys. Lett.* **1977**, *51*, 447–452.
- (20) Park, S. M.; Paffett, M. T.; Daub, G. H. *J. Am. Chem. Soc.* **1977**, *99*, 5393–5399.
- (21) Treibs, A.; Kreuzer, F. H. *Justus Liebigs Ann. Chem.* **1968**, *718*, 208–223.
- (22) Qin, W.; Baruah, M.; Van der Auweraer, M.; De Schryver, F. C.; Boens, N. *J. Phys. Chem. A* **2005**, *109*, 7371–7384.
- (23) Benniston, A. C.; Copley, G. *Phys. Chem. Chem. Phys.* **2009**, *11*, 4124–4131.
- (24) Bañuelos, J.; Arroyo-Córdoba, I. J.; Valois-Escamilla, I.; Alvarez-Hernández, A.; Peña-Cabrera, E.; Hu, R.; Zhong, Tang, B. Z.; Esnal, I. *RSC Adv.* **2011**, *1*, 677–684.
- (25) Loudet, A.; Burgess, K. *Chem. Rev.* **2007**, *107*, 4891–4932.
- (26) Ziessel, R.; Ulrich, G.; Harriman, A. *New J. Chem.* **2007**, *31*, 496–501.
- (27) Gomez-Duran, C. F. A.; Garcia-Moreno, I.; Costela, A.; Martin, V.; Sastre, R.; Banuelos, J.; Arbeloa, F. L.; Arbeloa, I. L.; Pena-Cabrera, E. *Chem. Commun.* **2010**, *46*, 5103–5105.
- (28) Ulrich, G.; Ziessel, R.; Harriman, A. *Angew. Chem., Int. Ed.* **2008**, *47*, 1184–1201.
- (29) Andrade, G. A.; Pistner, A. J.; Yap, G. P. A.; Lutterman, D. A.; Rosenthal, J. *ACS Catalysis* **2013**, *3*, 1685–1692.
- (30) Leonardi, M. J.; Topka, M. R.; Dinolfo, P. H. *Inorg. Chem.* **2012**, *51*, 13114–13122.
- (31) Khatchadourian, A.; Krumova, K.; Boridy, S.; Ngo, A. T. *Biochemistry* **2009**, *48*, 5658–5668.
- (32) Rosenthal, J.; Lippard, S. J. *J. Am. Chem. Soc.* **2010**, *132*, 5536–5537.
- (33) Royzen, M.; Wilson, J. J.; Lippard, S. J. *J. Inorg. Biochem.* **2013**, *118*, 162–170.
- (34) Nepomnyashchii, A. B.; Broring, M.; Ahrens, J.; Kruger, R.; Bard, A. J. *J. Phys. Chem. C* **2010**, *114*, 14453–14460.
- (35) Nepomnyashchii, A. B.; Bard, A. J. *Acc. Chem. Res.* **2012**, *45*, 1844–1853.
- (36) Venkatanarayanan, A.; Martin, A.; Keyes, T. E.; Forster, R. J. *Electrochem. Commun.* **2012**, *21*, 46–49.
- (37) Krumova, K.; Cosa, G. *J. Am. Chem. Soc.* **2006**, *132*, 17560–17569.
- (38) Galletta, M.; Puntoriero, F.; Campagna, S.; Chiorboli, C.; Quesada, M.; Goeb, S.; Ziessel, R. *J. Phys. Chem. A* **2006**, *110*, 4348–4358.
- (39) Lakshmi, V.; Ravikanth, M. *Chem. Phys. Lett.* **2013**, *564*, 93–97.
- (40) Nepomnyashchii, A. B.; Broring, M.; Ahrens, J.; Bard, A. J. *J. Am. Chem. Soc.* **2011**, *133*, 19498–19504.
- (41) Nepomnyashchii, A. B.; Broring, M.; Ahrens, J.; Bard, A. J. *J. Am. Chem. Soc.* **2011**, *133*, 8633–8645.
- (42) Nepomnyashchii, A. B.; Broring, M.; Ahrens, J.; Kruger, R.; Bard, A. J. *J. Phys. Chem. C* **2010**, *114*, 14453–14460.
- (43) Nepomnyashchii, A. B.; Cho, S.; Rossky, P. J.; Bard, A. J. *J. Am. Chem. Soc.* **2010**, *132*, 17550–17559.
- (44) Suk, J.; Omer, K. M.; Bura, T.; Ziessel, R.; Bard, A. J. *J. Phys. Chem. C* **2011**, *115*, 15361–15368.
- (45) Nepomnyashchii, A. B.; Pistner, A. J.; Bard, A. J.; Rosenthal, J. *J. Phys. Chem. C* **2013**, *117*, 5599–5609.
- (46) Rosenthal, J.; Nepomnyashchii, A. B.; Kozhukh, J.; Bard, A. J.; Lippard, S. J. *J. Phys. Chem. C* **2011**, *115*, 17993–18001.
- (47) Uhlich, N. A.; Sommer, P.; Bühr, C.; Schürch, S.; Reymond, J.-L.; Darbre, T. *Chem. Commun.* **2009**, 6237–6239.
- (48) Sahami, S.; Weaver, M. J. *J. Electroanal. Chem.* **1981**, *122*, 155–170.
- (49) Ammar, F.; Saveant, J. M. *J. Electroanal. Chem.* **1973**, *47*, 215–221.
- (50) Itaya, K.; Bard, A. J.; Szwarc, M. *Z. Phys. Chem. Neue. Fol.* **1978**, *112*, 1–9.
- (51) Bard, A. J.; Faulkner, L. R. *Electrochemical Methods: Fundamentals and Applications*; John Wiley: New York, 1980.
- (52) Lai, R. Y.; Bard, A. J. *J. Phys. Chem. B* **2003**, *107*, 5036–5042.
- (53) Arbeloa, T. L.; Arbeloa, F. L.; Arbeloa, I. L.; Garcia-Moreno, I.; Costela, A.; Sastre, R.; Amat-Guerri, F. *Chem. Phys. Lett.* **1999**, *299*, 315–321.
- (54) Ziessel, R.; Ulrich, G.; Harriman, A. *New J. Chem.* **2007**, *31*, 496–501.
- (55) Miao, W. In *Handbook of Electrochemistry*; Zoski, C. G., Ed.; Elsevier: Amsterdam, The Netherlands, 2007; p 541.
- (56) Sartin, M. M.; Camerel, F.; Ziessel, R.; Bard, A. J. *J. Phys. Chem. C* **2008**, *112*, 10833–10841.